SELECTIVITY VARIATION IN HYDROLYSIS OF PHENYL ACETATES BY SIMPLE MODIFICATIONS OF β -CYCLODEXTRIN.

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By reaction of β -cyclodextrin 6-monotosylate with alkyl mercaptans, 6-deoxy-6-alkylthio- β cyclodextrins, ζ , ζ , and ζ , were prepared. Studies of the hydrolyses of m- and p-substituted phenyl acetates showed that the well-known meta-selectivity effect occurred with 2 , while none was observed with A . This variation in selectivity was due to a change in the catalytic rate constant caused by the substituent on B-cyclodextrin.

In the past decade, many catalytic processes modeling enzymic reactions have been extensively studied by use of α - or β -cyclodextrins¹. Introduction of an appropriate catalytic group to the cyclodextrin has been successfully carried out to obtained a rather sophisticated model of an enzyme^{2}. Also, a dramatic enhancement of the inclusion ability of the cyclodextrin was attained by the capping of the cavity with an hydrophobic moiety³ or a metal complex⁴.

In the studies of the hydrolyses of substituted phenyl acetates by α - or β -cyclodcxtrins, meta-substituted phenyl esters were more rapidly hydrolyzed than the corresponding para-isomers, a phenomenon termed "meta-selectivity"^{1b}. This selectivity is apparently dependent on the depth of the cavity of the cyclodextrin and should be altered by appropriate modifications of the cyclodextrins. In our previous paper⁵, we demonstrated the reversion from "meta-selectivity" to "para-selectivity" by a complete capping of β -cyclodextrin: complete capping (1) decreased the maximum catalytic rate constant (k_c) for the para-esters to values quite similar to those for the corresponding mcta-isomers. Also, binding of the para-esters was enhanced to much greater degree.

In this paper, we wish to demonstrate that simpler modifications of β -cyclodextrin also change the selectivity for ester hydrolyses because a different mechanism come into play.

Modifications of β -cyclodextrin (2, 3 and 4) were carried out by the reaction of mercaptans and β -cyclodextrin 6-monotosylate, \mathcal{I} . The modified cyclodextrins were purified by repeated

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| $ACO-C6H4-X$ X | k_0 (x 10 ⁻³ sec ⁻¹) | | | K_A (x 10 ⁻³ M) | | |
|----------------------------------|--|---------------------|----------------------|-------------------------------|---------------------|-----------------|
| | | $\overset{3}{\sim}$ | $\stackrel{4}{\sim}$ | | $\overset{3}{\sim}$ | يمه |
| $p-NO_2$ | $90 \div 14$ | $106+1$ | $136 + 3$ | 5.1 ± 0.9 | $1.83 \div 0.03$ | 0.73 ± 0.03 |
| \underline{m} -NO ₂ | 521 ± 117 | $342 + 32$ | $150+5$ | $6.2 + 1.6$ | $3.1*0.2$ | 0.76 ± 0.04 |
| $p-Me$ | 4.91 ± 0.48 | 10.8 ± 2.3 | 24.4 ± 1.8 | 2.1 ± 0.3 | $1.5 + 0.5$ | 1.00 ± 0.12 |
| $m-Me$ | 18.5 ± 2.4 | 19.8 ± 0.4 | 21.1 ± 1.8 | 4.4 ± 0.7 | 2.62 ± 0.06 | 0.73 ± 0.12 |

Table I. Maximum Catalytic Rate Constants (k_c) and Dissociation Constants (k_d) for Reactions of Modified β -Cyclodextrins with m - and p-Substituted Phenyl Acetates^a.

^a In pH 10.60 (I=0.15) NaHCO₃-Na₂CO₃ buffer, 25°C, with 0.50-0.70% (v/v) CH₃CN added.

Table \Box . Selectivity in Reactions of Modified β -Cyclodextrins with m - and p -Substituted Phenyl Acetates (25°C. pH 10.60, 1=0.15).

| $ACO-C6H4-X$ Χ | k_c/K_d (M^{-1} sec ⁻¹) | | | (k_c/K_d) para $/(k_c/K_d)$ meta | | |
|------------------------|--|----------------------|---------------|------------------------------------|---------------|-------|
| | $\stackrel{2}{\sim}$ | $\stackrel{3}{\sim}$ | \mathcal{L} | | $\frac{3}{2}$ | يمه |
| $P-NO2$ | 17.9 | 58.1 | 187 | 0.213 | 0.520 | 0.940 |
| $\underline{m} - NO_2$ | 83.9 | 111.7 | 199 | | | |
| $p-Me$ | 2.35 | 7.11 | 24.4 | 0.564 | 0.940 | 0.841 |
| m-Me | 4.17 | 7.56 | 29.0 | | | |

recrystallization from water and characterized by IR, NMR, FD mass spectrometry, and elemental analyses⁷. The hydrolyses (pH 10.60) of meta- and para-nitro- and methyl-phenyl acetates were measured spectroscopically at 25° in the presence of an excess of the modified or native β -cyclodextrins and showed pseudo-first-order kinetics $^8.$ From the Eadie treatment of the kinetic data⁸, the dissociation constants (K_d) and the maximum catalytic rate constants (k_c) were obtained (Table I and Scheme I).

As' the hydrophobic nature of the substituent of the cyclodextrin increased from2 to 4, the following results were obtained; (1) the selectivity factor, (k_c/K_d) increased (Table II), (2) the ratio of the selectivity factor, $(k_c/K_d)_{\text{para}}/(k_c/K_d)_{\text{meta}}$, increased up to almost unity, (3) all values of K_d decreased, (4) the K_d value for the meta-ester was always similar to that for the corresponding para-ester, and (5) the value of k_c for the para-esters increased while that for the meta-esters decreased or increased only slightly. While the well-known "meta-selectivity" effect was highly conspicuous with 2 , essentially no selectivity was observed with 4 . This lack of selectivity with A was due primarily to an increase or decrease of k_c values for the para- or meta-esters, respectively.

Thus the well-known "meta-selectivity" effect is not a general property of cyclodextrin catalysis. Instead, selectivity can be easily varied by simple modifications of the cyclodextrin structure which in turn leads to changes in k_c and K_d depending on the functional group utilized in the modification.

Modified Cyclodextrin + Phenyl Acetate $\frac{1}{\sqrt{2}}$ Com $^{\sim}$ 2 \int_0^{∞}

Acetyl-Modified Cyclodextrin + Phenolate

where
$$
K_d = \frac{k_c + k_2}{k_1}
$$

$$
\hbox{Scheme} \quad \hbox{I}
$$

References and Notes

1. For examples; (a) F. Cramer and W. Kampe, J. Amer. Chem. Soc., 87, 1151 (1975). (b) R. L. van Etten, J. F. Sebastian, G. A. Clowes, and M. L. Bender, ibid., 89, 3242 (1967). (c) R. Bleslow and P. Cambell, Bioorg. Chem., 1, 140 (1971). (d) I. Tabushi, K. Fujita, and

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- 4. I. Tabushi, N. Shimizu, T. Sugimoto, M. Shiozuka, and K. Yamamura, J. Amer. Chem. Sot., 99, 7100 (1977).
- 5. K. Fujita, A. Shinoda, and T. Imoto, J. Amer. Chem. Soc., 102, in press (1980).
- 6. The procedure of the present modification was similar to that described elsewhere. See ref.(3c).
- 7. Molecular weights of 2, 3, and 4 were determined by means of the field desorption mass spectra which will be reported in near future. 2: Found: C, 40.76, H, 6.49. Calcd for $C_{AA}H_{7A}O_{35}$. 6H₂0: C, 40.55, H, 6.65. ¹H-NMR (D₂O): 62.62 (2H, SCH₂CH₂OH), 2.9 (2H, cyclodextrin-CH₂S), 4.90 (7H, C₁H of cyclodextrin) and 3.2-4.1 (42H, SCH₂CH₂OH and cyclodextrin protons other than C₁H). $\frac{3}{2}$: Found: C, 44.61, H, 6.46. Calcd for C₄₅H₇₆O₃₄S'H₂O: C, 44.63, H, 6.49. ¹H-NMR (D₂O): 60.83 (3H, CH₂CH₂CH₃), 1.45 (2H, CH₂CH₂CH₃), 2.46 (2H, <u>CH₂C₂H₅)</u>, 2.9 (2H, cyclodextrin-CH₂S), 4.90 (7H, C₁H of cyclodextrin), and 3.2-4.1 (40H, cyclodextrin protons other than C_1H). $4:$ Found: C, 45.58, H, 6.59. Calcd for $C_{47}H_{80}O_{34}S\cdot H_2O$: C, 45.55, H, 6.67. 1 H-NMR (D₂O-DMSO-d₆): 60.80 (9H, CMe₃), 2.66 (2H, SCH₂CMe₃), 2.9 (2H,cyclodextrin- CH_2S), 4.84 (7H, C,H of cyclodextrin), and 3.2-4.1 (40H, cyclodextrin protons other than C₁H). Ir spectra of 2, 3 and 4 were very similar to that of B-cyclodextrin.
- 8. Measurement and treatment of the kinetic data were carried out as described elsewhere. See ref. $(1b)$.

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